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UNIVERSITY OF  
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## ANIMAL CARE SERVICES NEWSLETTER

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### Renovations and Equipment Improvements

The cage wash, Infectious Isolation Unit, and SPF rodent housing facilities within the Communicore building are operational. Additional minor improvements in the cage wash area and SPF area are planned. The new cat housing facility at the Bivens Arm Animal Research Facility (BAARF) is moving ahead. We plan on having this building operational around June 2003.

### New Policies

**NOTE:** ACS policies are located on the web at <http://www.health.ufl.edu/acs>

#### HSC Animal Transportation Policy

All animal transportation between and within buildings at the University of Florida Health Science Center must conform to this policy. This policy describes:

- Movement of animals within a building at the University of Florida
- Movement of animals between buildings at the University of Florida
- Transportation of live or dead animals containing radioactive isotopes
- Transportation of animals treated with human pathogens or carcinogenic material
- Species specific guidelines

Please, visit the ACS web site to download this policy ([http://www.health.ufl.edu/acs/policy/Transport+Policy+2\\_1\\_03.doc](http://www.health.ufl.edu/acs/policy/Transport+Policy+2_1_03.doc)).

Effective date of this policy – February 1, 2003

#### Transfer of Animals between Rooms Policy

Movement of animals from one room to another is the sole function of ACS staff. Animal movements are coordinated through the Clinical Veterinarian who knows the pathogen status in each room. To avoid pathogen spread within the facility, research staff should not move animals between rooms.

To move animals between rooms, please go to <http://www.health.ufl.edu/acs/forms/relocation.doc> to download the *Animal Relocation Request Form*. This form must be completed and submitted to the front office (CB 160) via email or fax (392-3766) two business days prior to the requested moving date. Forms dropped in the facility drop box will require four business days for processing. Rushed orders will be assessed an administrative fee.

Effective date of this policy – March 1, 2003

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Animal Care Services (ACS) and the office of the Attending Veterinarian publishes, every other month, a newsletter to communicate with those who use or provide support to the animal care program at the University of Florida. This newsletter contains new policies, new services provided, updates on animal facility improvements, updates on federal regulations, policies and other items of importance to animal users. We use e-mail as our primary method of distributing the newsletter and also have copies of the newsletter available in the animal facilities. Please print this newsletter and distribute it to members of your department. If you did not receive this newsletter by email, and would like to be added to the email list, please send an email message to [tmkegley@vpha.health.ufl.edu](mailto:tmkegley@vpha.health.ufl.edu) with "add to ACS newsletter" under subject. All issues will be maintained on our website.

## Policy on Requesting Clean Rodent Cages

Investigators may need to have rodent cages available for separating weanling cages, replace wet cages, or for other appropriate reasons. Animal Care Services will provide clean rodent cages with bedding and other caging accessories for these purposes and will deliver these items to a designated animal room. We prefer to deliver this equipment to the room rather than have the investigator come to our clean cage storage rooms located in the Communicore and MBI. This practice is intended to reduce the risk of spreading pathogens, such as pinworms, between animal rooms. It is possible to carry dust particles holding pathogens on clothing and transfer pathogens to other clean cages. If you need clean cages, please complete a *Husbandry Special Request Form* located at <http://www.health.ufl.edu/acs/formsorder.htm>. In addition, we would also appreciate investigators placing their requests on the appropriate dry erase boards located outside the clean side of the cage wash facilities in the Communicore, MBI animal facilities, or in the Communicore SPF facility. If you need this service on more than one occasion, only one requisition needs to be completed with start and end dates, day and time of the week, and other specifics. We will begin to keep a small number of clean cages, kept in a plastic bag, or on the ventilated racks within each animal room to be used as needed, however please submit a requisition to ensure that you have an adequate number of cages and accessories. If you have any questions regarding this policy, please contact Mike Riley at 392-4484, <mailto:mpriley@vpha.health.ufl.edu>.

Effective date of this policy – March 1, 2003

## Animal Use Protocols & ACS Veterinary Consultations

Effective October 3, 2002, Animal Care Services assumed administrative control over the protocol consultation process. All protocols are sent to [vconsult@grove.ufl.edu](mailto:vconsult@grove.ufl.edu). From there, ACS begins the consultation process and tracks each protocol from the day it is e-mailed to vconsult to the day it is submitted to IACUC. The average consultation review time since October 3 has been ten days. Some protocols take longer than usual because they require additional attention or outside consultation review.

## Notice to All Investigators Using Rodents at the Communicore Building Regarding Pinworm Infestation

### Background

Some animal rooms in conventional housing tested positive for pinworms during our routine January 2003 checks. To control this problem effectively, all conventional rodents have been placed on medicated diet (with fenbendazole) for six weeks since the week of February 10. Alternative treatment is ivermectin in drinking water if for some reason fenbendazole can not be used in your research animals. If you have any question or concerns or need special arrangements, please contact Dr. Sunder Shrestha (846-0981 [sshres@vpha.health.ufl.edu](mailto:sshres@vpha.health.ufl.edu)).

Recent renovations, non-availability of autoclaves, cage washer installation and movement of animals related to the renovations may have contributed to the spread of pinworms. We need help from all concerned in eliminating and controlling this problem.

### MANDATORY Pinworm Decontamination

Goal: To prevent the perpetuation and spread of pinworms through contact of treated animals with viable pinworm eggs. This requires environmental clean-up to remove any pinworm eggs present from previously shedding animals and treatment of at-risk animals to prevent reinfection until environmental decontamination is completed.

Timing: Rodents in all conventional rooms are being treated for pinworms with fenbendazole-medicated feed for nine weeks. After this time animals should not be shedding pinworm eggs. Decontamination procedures will follow in the animal room (includes room decontamination and changeover of all cages, racks, feeders, etc.) as well as the completion of all decontamination in any lab areas. If any pinworm eggs are detected during the nine-week follow-up testing, treatment and environmental decontamination will be repeated.

ACS will perform all decontamination in the animal facilities, but investigators are responsible for decontamination of their own lab areas, carts, and research equipment. This should be performed as soon and as carefully as possible.

- A delay in cleaning up your lab will delay the time when your animals can come off treated feed.
- Insufficient decontamination may result in reinfection of your and other PI animals.

### **Pinworm Decontamination Methods**

Pinworm eggs are generally considered resistant to disinfectants and desiccation. They are also lightweight and sticky, and can be transferred on clothing, objects, and dust particles. The primary method of eliminating pinworm eggs (short of autoclaving) is by PHYSICAL REMOVAL. Wear disposable gloves, gowns and shoe covers while performing decontamination procedures.

Autoclavable items: Autoclaving using a standard sterilization cycle reliably kills pinworm eggs.

Sturdy, heatproof plastic, glass, or metal items without excessive "dead space" (hidden corners, etc.): Cage washing adequately removes eggs through a combination of water temperature, pressure, detergent, and dilution.

Washable walls, ceilings, floors, windows, etc.: Use of a pressure washer accompanied by scrubbing with a brush adequately removes eggs through a combination of water pressure, detergent, physical dislodgment, and dilution. This method will be used to clean animal rooms.

Countertops, less sturdy or complex items: Hand washing with detergent/soap and water, accompanied by scrubbing and copious rinsing, removes eggs through physical dislodgment and dilution.

Clothing: Machine laundering adequately removes eggs through a combination of water temperature, agitation, detergent, and dilution.

Non-washable items (such as books and notebooks): Wipe clean with damp paper towels if possible. Replace with new items, storing old items away from animal use areas.

Chemical sterilization (such as formaldehyde gas) is effective but may be complicated and is hazardous to personnel. Please, contact EH&S prior to using this method.

After detergent cleanup, a follow-up treatment with quaternary ammonium compounds (MB-10, Clidox, Roccal-D) and bleach may be helpful. Be aware that ammonia fumes are noxious and that ammonia-containing compounds must never be used in conjunction with bleach. Please, contact EH&S prior to using this method.

### **To Contain this Infestation, We Request All Research Staff to Follow these Procedures:**

- Visit your clean rooms prior to the infected room. Do not back track from the infected room to your clean rooms.
- Strictly follow all PPE requirements listed on the room's door. Remove the PPE as you exit the room. Wearing it outside the room only spreads the eggs.
- If you need to transport your animals out of the room, they must be covered with a filter top and the cages sprayed with MB-10 (chlorine dioxide).
- If you use the animals in a procedure room or in your lab you must:
  - Use animals from your clean rooms first, disinfecting between groups.
  - Sanitize the area where the animals are used, equipment used including transport carts, and wear a clean lab coat when handling the animals.
- If you remove cages to the soiled cage wash, cover the cages with a filter top and spray the cages with MB-10.
- If you need to get clean cages and equipment to separate or transport your animals in this room, please get the clean items prior to entering the room.

## **Infectious Isolation Unit (IIU) Required Training**

The IIU located in the HSC Animal Care Services facility is now operational. Investigators whose animals are housed in the IIU will be assessed a 20% surcharge over and above the SPF per diem rate as posted on the ACS web site. For more information, please check our web site at <http://www.health.ufl.edu/acs/index.htm> under "Services and Fees."

Beginning March 17<sup>th</sup> the doors to the IIU will be locked down. To obtain access, research staff must first complete the IIU auto tutorial (see ACS web site, <http://www.health.ufl.edu/acs/index.htm>), followed by a facility practicum/tour. For the practicum/tour, please call the ACS front office at 392-2977 to schedule a session. After successful completion of the auto tutorial and facility practicum, you may request facility access to the IIU by submitting a *Request for Access to Animal Facilities Form* (see ACS web site at <http://www.health.ufl.edu/acs/formsorder.htm> and click on *Forms and Ordering*).

If you have any questions regarding the auto tutorial or facility tour, please contact Kelly Flint at 846-2416, [kflint@vpha.health.ufl.edu](mailto:kflint@vpha.health.ufl.edu).

## **Specific Pathogen Free Facility (SPF) Required Training**

To obtain access to the HSC SPF facility, research staff must participate in a facility practicum/tour by contacting the ACS front office at 392-2977 to schedule a session.

If you have any questions regarding the facility tour, please contact Travis Cossette at 846-0982, [travisc@ufl.edu](mailto:travisc@ufl.edu).

## **ACS Staff Announcements and AALAS Training Classes**

Mr. Luis Zorrilla (Surgery Manager) received AALAS certification at the Laboratory Animal Technician (LAT) in December 2002. This is the second of four AALAS certification levels, which attests to a high level of technical proficiency.

Ms. Kate Gilleece (Animal Technician) and Ms. Georgette Dee (Animal Technician) received AALAS certification at the Assistant Laboratory Animal Technician (LAT) in December 2002. This is the first of four AALAS certification levels, which also attests to a high level of technical proficiency.

Beginning February 18, 2002, ACS is conducting AALAS training classes at the ALAT and LAT level. Anyone interested in attending these classes may contact Luis Zorrilla at 392-9948, [zorro21@vpha.health.ufl.edu](mailto:zorro21@vpha.health.ufl.edu). Among the topics presented are husbandry, animal care, species-specific needs, regulations, and others.

## **Miscellaneous**

### **Useful Web Sites**

The ACS web site has a number of useful links for investigators and their staff. Please take advantage of these links.

### **ACS Security Problems**

As many of you know, the security system in the Communicore Animal Care Facility has not been working for over the past three weeks. The doors remain locked, but cannot be opened with a "swipe card" except at the loading dock. Until we get this resolved, we will leave the entrances to the facility open between 7:00 and 5:00 PM Monday through Friday and the West entrance to the Communicore Animal Facility (entrance down from the Sun Terrace Cafeteria) will be opened from 7:00 - 11:00 am Saturday and Sunday. If you need access after hours, you will need to contact campus security at 392-1111.

We have arranged through Siemens Corporation to have three biometric readers installed in the Communicore building, which is consistent with our long range plan. Because there are considerable costs and delays associated with repairing the existing system, we would rather invest in the new technology. We expect to have the new system functional within the next three weeks.

